

# **Modeling Inactivation Kinetics for *Enterococcus faecium* on the Surface of Peanuts during Convective Dry Roasting**

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IFTPS Charles Stumbo Paper Winner

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# Salmonella thermal resistance

- Foodborne illness risk can be decreased through thermal processing steps (e.g., roasting, baking, toasting, extruding)
- *Salmonella* is notoriously hard to kill in low-moisture environments<sup>1,2,3</sup>
- Decreased water activity (or moisture) is generally correlated with a higher D-value and z-value<sup>4</sup>
- FSMA requires a validated hazard reduction<sup>5</sup>

From Smith et al. (2016):

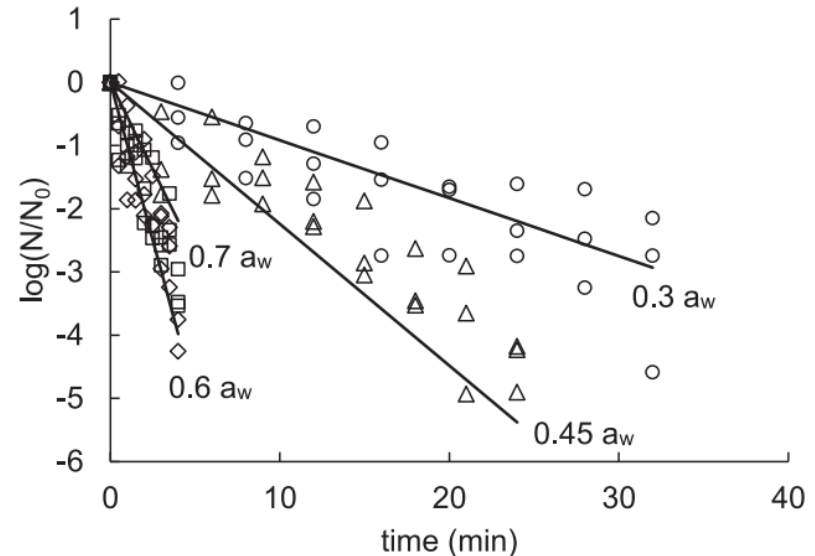


FIGURE 2. Survivor curves for *Salmonella* inoculated into wheat flour at four nominal  $a_w$  levels and treated at 80°C. Lines are log-linear/modified-Bigelow model (equations 1 and 6) predictions.

# Model-based validations: a historical perspective

- Conduct a temperature distribution study to get an idea of thermal distribution in a retort (i.e., where is the coldest spot?)<sup>6</sup>
- Conduct a thermal penetration study to determine the rate at which the slowest heating can will heat up (i.e., what is the minimum time required to achieve specified 12D lethality?)<sup>6</sup>
- Heating food in a can or pouch inside a retort is predictable
- Death kinetics are consistent and characterized based on temperature
- With well-characterized product heating, we can use models to predict lethality and validate a retort

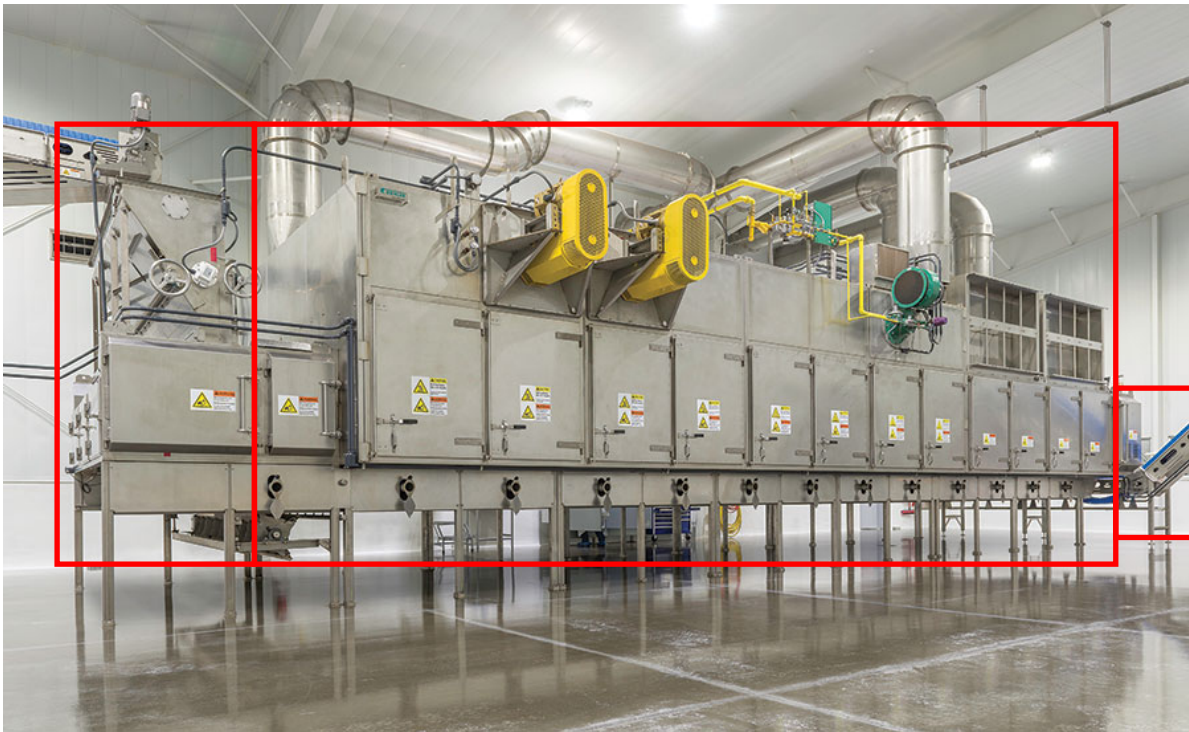


<https://funfactz.com/food-and-drink-facts/canned-food-safe-to-eat/>

# Model-based validations: a historical perspective

- Once we know where the lowest lethality is achieved, the log reductions can be calculated using general method<sup>7</sup>
- Numerical integration of D- and z-value models as a function of process time and product temperature<sup>7</sup>
  - Graph lethal rate,  $10^{\frac{T-T_{ref}}{z}}$  [=] log reduction/time
  - Calculate area under curve to get lethality [=] log reductions
- Kinetics for *C. botulinum* (target pathogen) are well-known

# Digression for a quick roaster tutorial

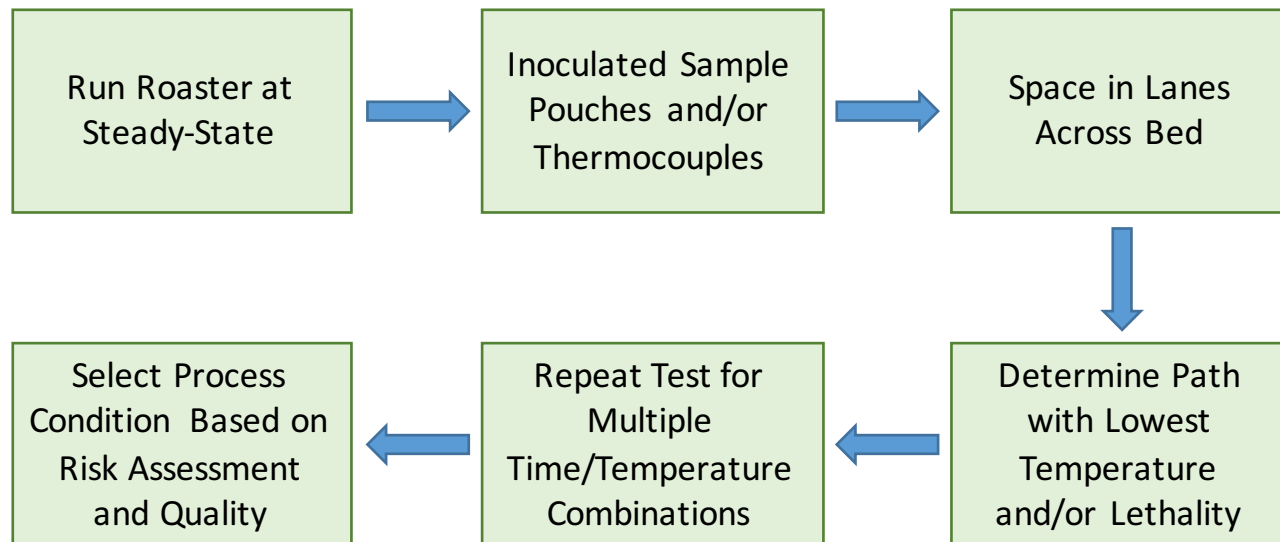


<https://www.plantengineering.com/single-article/advances-in-dryer-design/d2a11a53b2316f5ae3f602c631acdafd.html>

- Infeed (hopper)
- Preheat
- 4 heating zones, individually temperature controlled
- Cooling
- Discharge

# State-of-the-art for roaster validation

- How do we validate flat-bed dry roasters?<sup>8</sup>

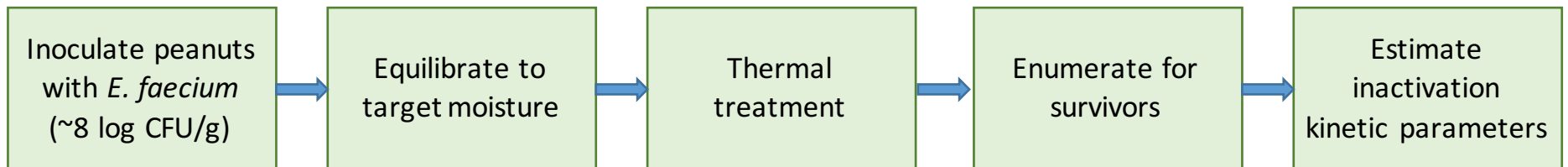


Wouldn't it be great if we could use a model and not have to worry about inoculated samples?

## Why use a model?

- Run “what-if” scenarios to change a process or if there is a process deviation<sup>9</sup>
- Temperature (and/or moisture, humidity, velocity, etc.) mapping is easier than a microbial validation
- Reducing sources of variability

# Experimental Overview





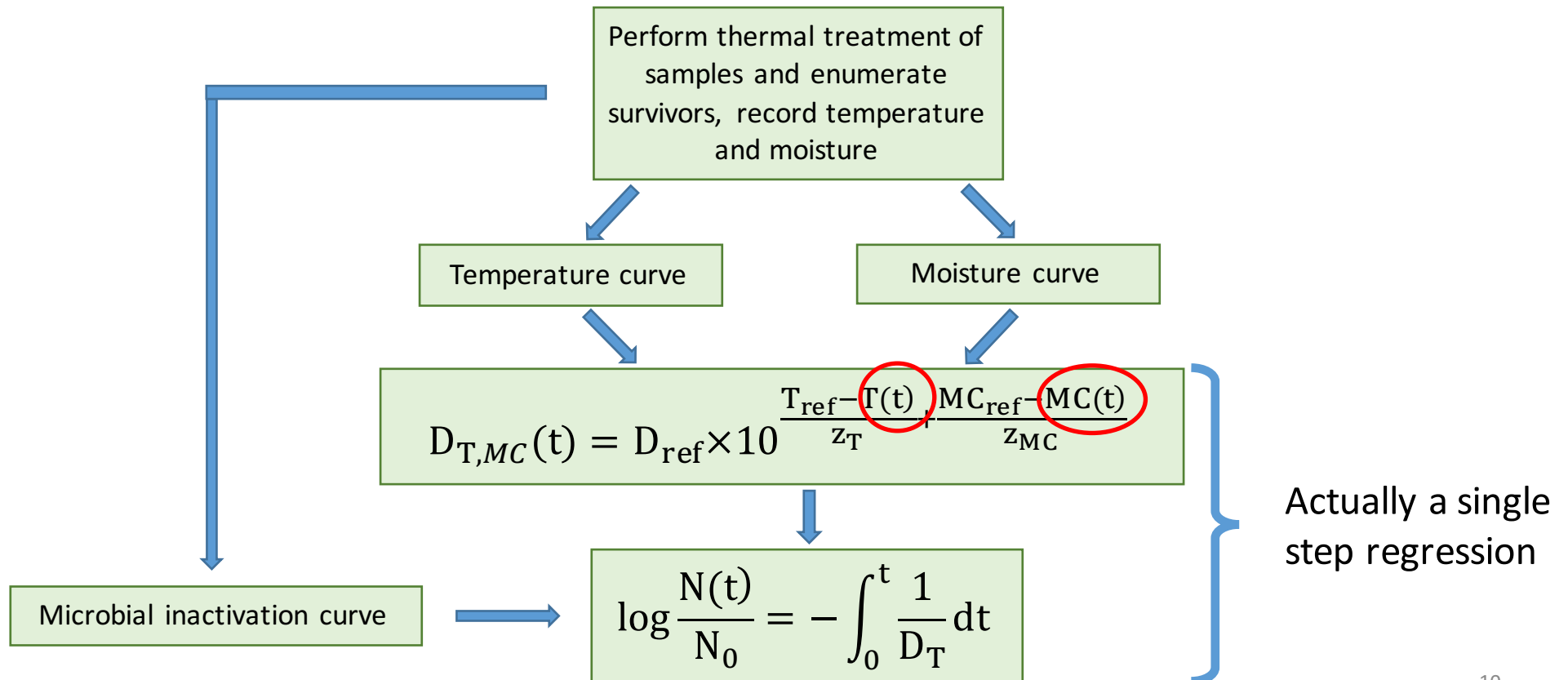
# Thermal Treatment

- Roasting conditions:

	1.0 m/s	1.3 m/s
121°C	x3	x3
149°C	x3	x3
177°C	x3	x3



# Estimate Inactivation kinetics



## Models Used

$$\log \frac{N(t)}{N_0} = -\frac{t}{D}$$

$$\log \frac{N(t)}{N_0} = -\left(\frac{t}{D}\right)^n$$

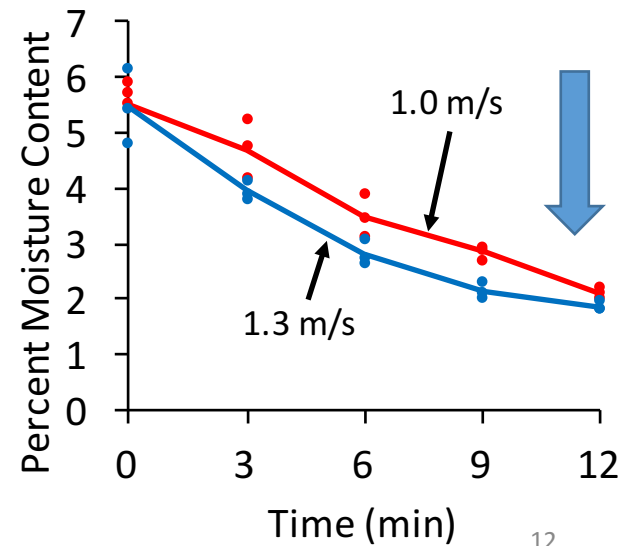
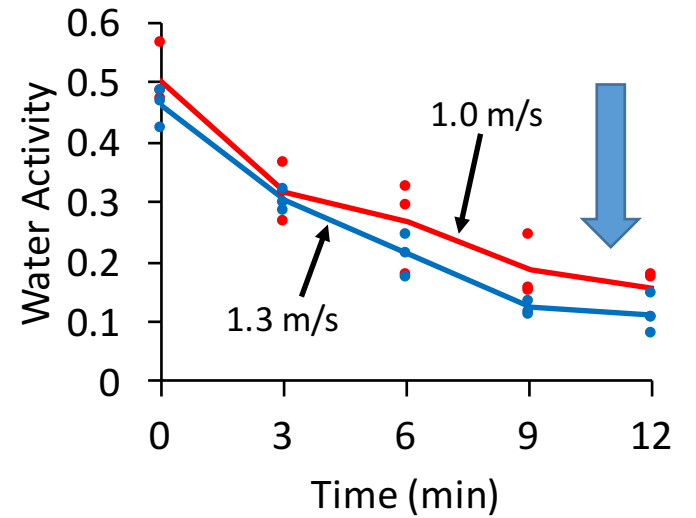
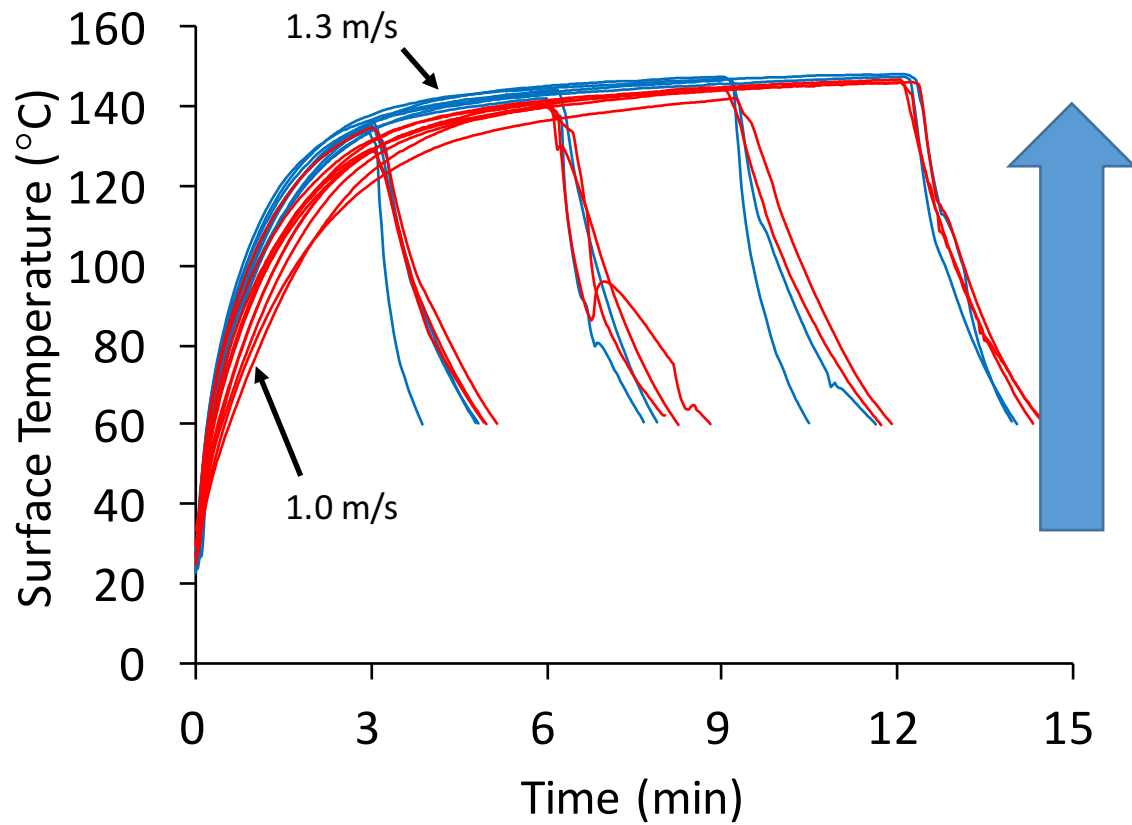
(yes, that's actually  $\delta$ !)

$$\longrightarrow D(T) = 10^{f(T)} = 10^{\frac{T_{ref}-T}{z_T}}$$

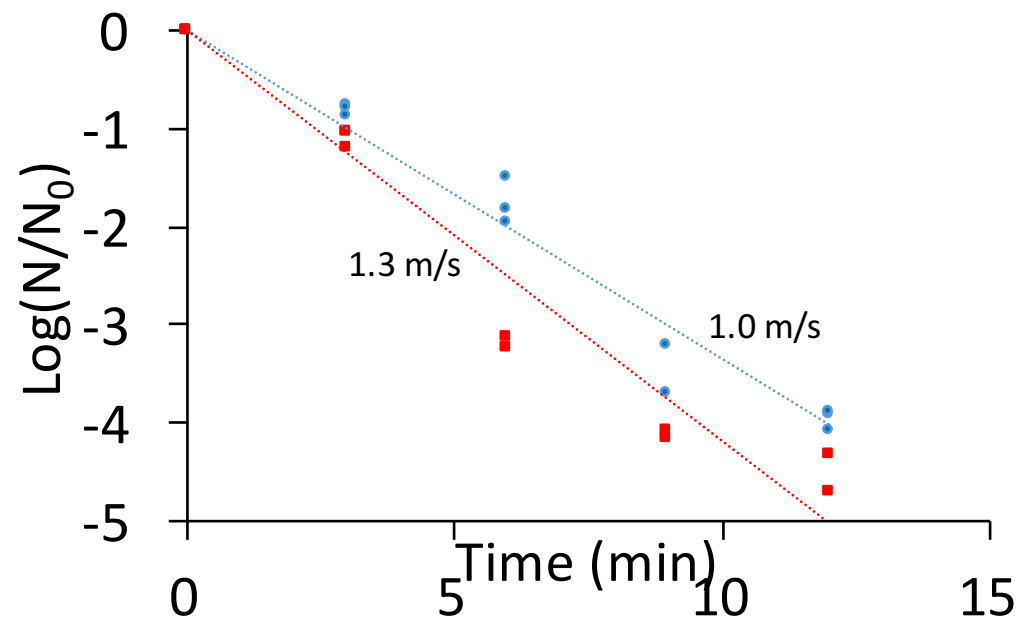
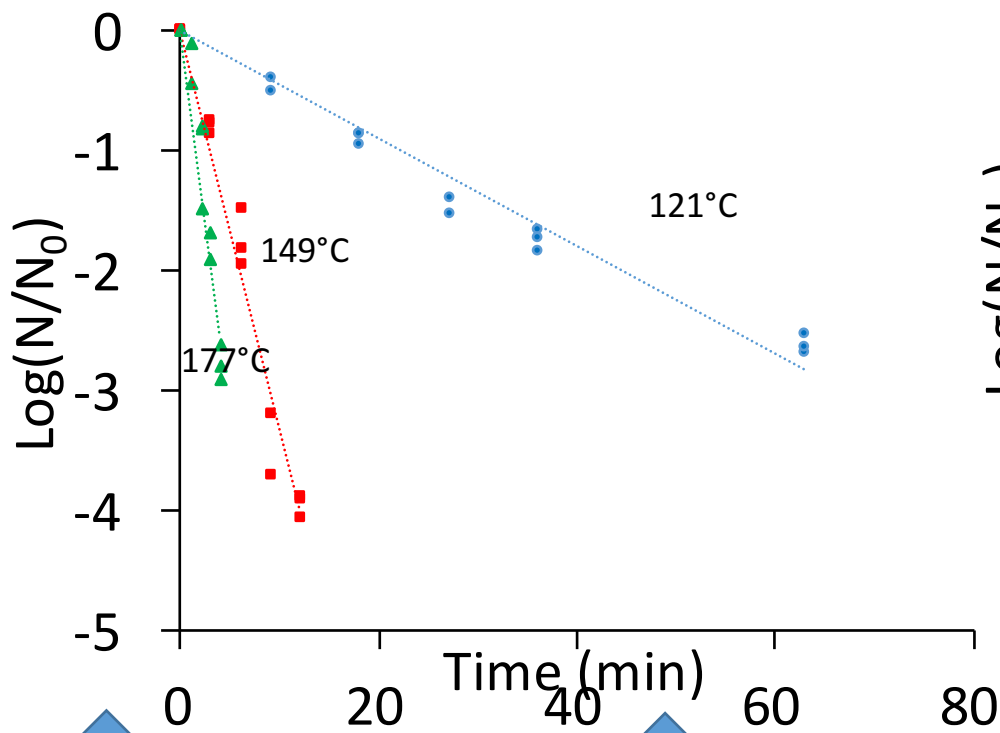
$$\longrightarrow D(T, a_w) = 10^{f(T)+f(a_w)} = 10^{\frac{T_{ref}-T}{z_T} + \frac{a_{w,ref}-a_w}{z_{a_w}}}$$

$$\longrightarrow D(T, \%MC) = 10^{f(T)+f(\%MC)} = 10^{\frac{T_{ref}-T}{z_T} + \frac{MC_{ref}-MC}{z_{MC}}}$$

# Results – Physical Results



# Results – Microbiological Results



**Temperature**



**Air Velocity**



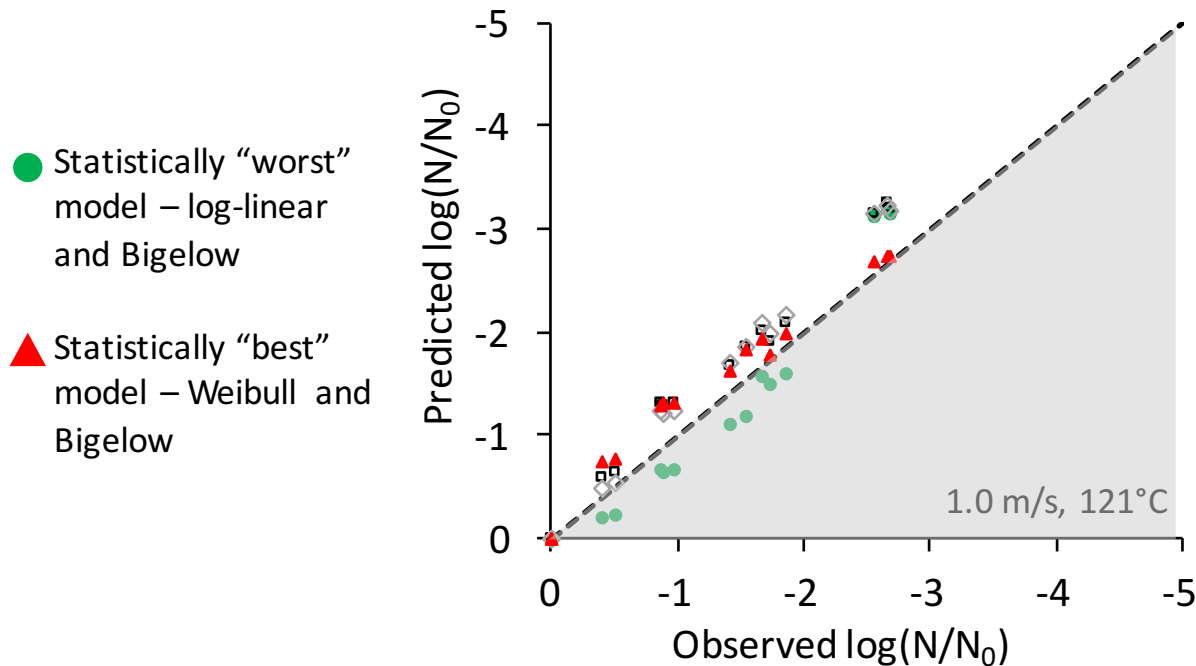
**Microbial Inactivation**

# Results – Parameter Estimates

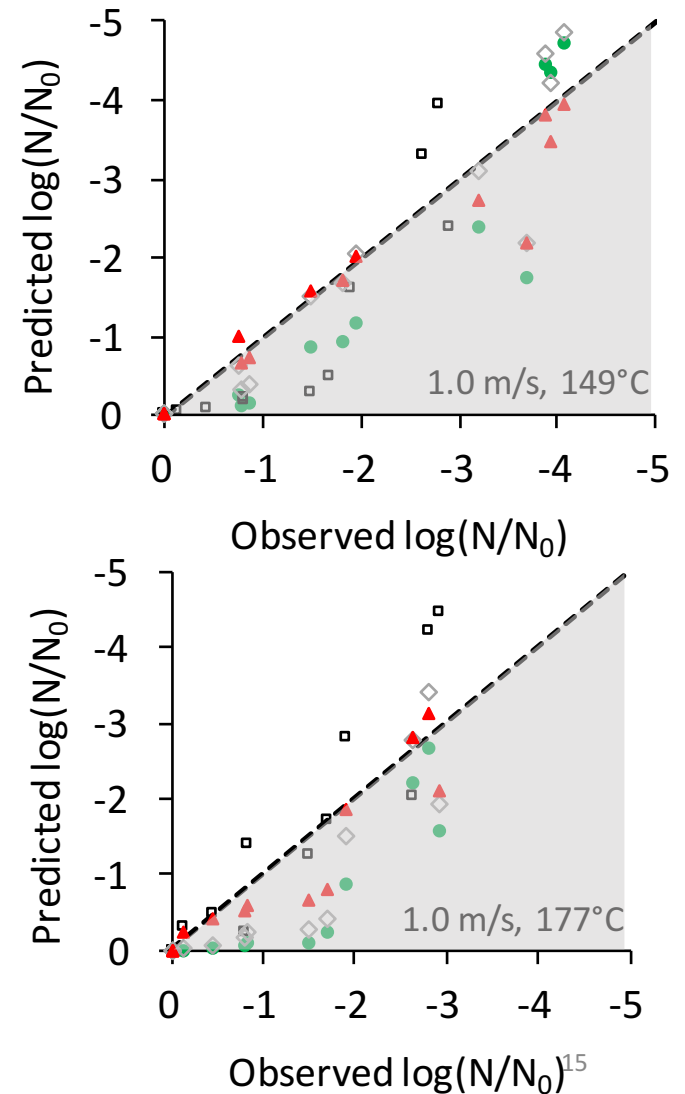
Models Used		Optimized Reference Conditions	Parameter Information			Model Statistics			
Primary	Secondary		Name	Value	Relative Error (%)	Mean Residuals	RMSE	Maximum Correlation	AIC <sub>c</sub>
Log-Linear	Bigelow	$T_{ref}=139^{\circ}\text{C}$	$D_{ref}$ (min)	3.04	3.71	-0.307	0.674	-0.0367	-64.8
			$z_T$ ( $^{\circ}\text{C}$ )	22.7	3.03				
Log-Linear	Modified Bigelow ( $a_w$ )	$T_{ref}=138^{\circ}\text{C}$ $a_{w,ref}=0.236$	$D_{ref}$ (min)	2.94	2.87	-0.159	0.534	-0.425	-103
			$z_T$ ( $^{\circ}\text{C}$ )	25.8	2.9				
			$z_{aw}$	0.532	13.8				
Log-Linear	Modified Bigelow (%MC)	$T_{ref}=138^{\circ}\text{C}$ $MC_{ref}=3.33\% \text{ MC, db}$	$D_{ref}$ (min)	2.97	2.86	-0.156	0.532	-0.3233	-104
			$z_T$ ( $^{\circ}\text{C}$ )	24.9	2.71				
			$z_{MC}$ (%MC, db)	5.01	14.0				
Weibull	Bigelow	$T_{ref}=132^{\circ}\text{C}$	$\delta_{ref}$ (min)	1.66E-06	9.11	-0.0188	0.379	0.1245	-163
			$z_T$ ( $^{\circ}\text{C}$ )	8.75	2.87				
			n	0.22	0.38				



# Results – Model Biases



Statistical differences were clear,  
but practical differences were minor



# References

1. Archer, J., E. T. Jervis, J. Bird, and J. E. Gaze. 1998. Heat Resistance of *Salmonella weltevreden* in Low-Moisture Environments. *J. Food Prot.* 61:969–973.
2. Casulli, K. E. 2016. Improving Pathogen-Reduction Validation Methods for Pistachio Processing. Master of Science Thesis in Biosystems Engineering, Michigan State University.
3. Stasiewicz, M. J., B. P. Marks, A. Orta-Ramirez, and D. M. Smith. 2008. Modeling the effect of prior sublethal thermal history on the thermal inactivation rate of *Salmonella* in ground turkey. *J. Food Prot.* 71:279–285.
4. Smith, D. F., I. M. Hildebrandt, K. E. Casulli, K. D. Dolan, and B. P. Marks. 2016. Modeling the Effect of Temperature and Water Activity on the Thermal Resistance of *Salmonella* Enteritidis PT30 in Wheat Flour. *J. Food Prot.* 79:2058–2065.
5. US Food and Drug Administration. 2018. Food Safety Modernization Act Final Rule for Preventive Controls for Human Food. Center for Food Safety and Applied Nutrition. <https://www.fda.gov/food/guidanceregulation/fsma/ucm334115>. Accessed 13 Jan 2019.
6. Institute for Thermal Processing Studies. 2014. Guidelines for Conducting Thermal Processing Studies. <http://iftps.org/wp-content/uploads/2017/12/Retort-Processing-Guidelines-02-13-14.pdf>. Accessed 13 Jan 2019.
7. Casulli, K. E., S. Calhoun, and D. W. Schaffner. 2019. Modeling the risk of salmonellosis from consumption of peanuts in the United States. *J. Food Prot.* Accepted.
8. Anderson, D., and L. A. Lucore. 2012. Validating the Reduction of *Salmonella* and Other Pathogens in Heat Processed Low-Moisture Foods. *Alliance for Innovation and Operational Excellence, Alexandria, VA*.
9. Dolan, K. D., and D. K. Mishra. 2013. Parameter estimation in food science. *Annu. Rev. Food Sci. Technol.* 4:401–422.



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# Questions/Comments

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